

ROLE OF PROTEINS IN RELATION TO DIABETES MELLITUS

Pushpa Durgawale, Priya Lanjekar and B.D. Punekar

Though the mechanism of metabolism of carbohydrate and protein is different, they take place simultaneously in the human system and are closely integrated to each other. It is seen that protein malnutrition in young adulthood leads to a malnutrition related diabetic syndrome in many developing countries.¹ The possibility that malnutrition in early infancy and childhood can result in partial failure of the β -cell function and clinical onset of diabetes later in life deserves further study.²

With experimental models of protein malnutrition it has been possible to reproduce the impaired glucose tolerance and decreased insulin secretory response to glucose in human Kwashiorkor.³⁻⁷ *In vitro* experiments on rats show that insufficient protein diet for a limited period results in diminished insulin secretory response to glucose after the rats are returned to an adequate diet.

That protein malnutrition affects the insulin secretion and glucose tolerance has been shown by Ingemar Swenne and Workers⁸ on experimental animals of same age and sex divided into two groups: one fed with low protein diet and the other fed the normal diet for a limited period of time. Subsequently, when both the groups were treated with a diet of high nutritional efficiency, it was seen that there was no change in weight gain and growth in protein malnourished rats compared to normal rats. However, in malnourished rats impairment of glucose tolerance and an increased hypoglycemic response to exogenous insulin was seen. These observations indicate that the impairment of glucose tolerance in protein malnutrition is caused by deficient insulin secretion rather than by changes in peripheral insulin action. On a diet of higher nutritional efficiency, the serum insulin levels of malnourished rats remain lower than the normal rats indicating a functional impairment of pancreatic islets due to protein malnutrition which is also irreversible.

When different tissue samples of normal and protein malnourished rats were analysed⁹, it was observed that malnutrition resulted in decrease in pancreatic size, diameter and volume of pancreatic islets. It was also observed⁹ that stimulation of pancreas with glucose load as well as with synthetic stimulators like A 23187 (Calcium ionophore which elicits first phase of insulin secretion)

Department of Biochemistry Krishna Institute of Medical Sciences, Karad 415110, Maharashtra

and TPA (4-B-phorbol-12 myristate-13 acetate which stimulates the second phase of insulin secretion) resulted in insulin secretion in both the groups. When insulin secretion was expressed as MU insulin/ml/mg. dry pancreatic weight, it was observed^{5,8,7} that, there was no significant difference in insulin secretory capacity of pancreas in low protein and normal-fed rats. Since the pancreatic size was decreased in protein malnutrition it resulted in proportionately lesser insulin secretion.

Different tissue samples of both the groups were analysed for DNA^{10,11} and protein¹² content and Protein/DNA ratio was used as an index of cell size. It was observed that in normal rats Protein/DNA ratio was increased with age, whereas, in protein malnourished rats the ratio was lower compared to normal rats of the same age. Even on treatment with a diet of higher nutritional efficiency in later life, the ratio remained lower in protein malnourished rats compared to normal rats. Persistent reduction of Protein/DNA ratio in malnourished animals and humans¹³ indicates that despite the treatment with an adequate diet, the increase in cellular proteins and growth in size are impaired as insulin stimulates protein biosynthesis and cytoplasmic growth rather than cell replication^{14,15}.

It is observed that in human Kwashiorkor¹⁶ and experimental protein malnutrition^{17,18,19} there is atrophy and disruption of normal pancreatic morphology and reduction in islet number, total islet mass²⁰ and size of individual β -cells.¹⁸ Possibly, in such animals normal pancreatic cell size and cell number required to maintain an adequate insulin secretion is not attained and as a result they are unable to respond to diabetogenic stimuli and nutritional challenges and are vulnerable to diabetes in later life.

While planning a diet of diabetic patient, the natural starch-protein interaction also may be taken into consideration. A study conducted by Rao and co-workers²¹ showed that various factors influence the rate of digestion of starch such as the nature of starch²², the natural starch-protein interaction,²³ the presence of fibres and anti-nutrients such as lectins, phytates and enzyme inhibitors^{24,25}. Nearly 10 to 20% of the starch in wheat flour is malabsorbed and it was interesting to find that²³ removing gluten from the wheat flour eliminated the malabsorption and subsequently adding back gluten to the gluten-free wheat flour did not reverse the effect. These observations raised the question whether the natural starch-protein interaction is responsible for the reduced digestibility of starch and if so what are its implications in terms of gastro-intestinal physiology in malabsorption states like celiac disease²⁶ which is usually associated with diabetes.

In vitro experiments by Rao²¹ showed that the natural starch-protein interaction decreased the starch-digestion products in dialysate. In their experiments, white bread, gluten-free bread and gluten-free bread+gluten (externally added) were used. They observed that concentration of starch digestion products was significantly low for white bread compared to gluten-free bread and subsequent addition of gluten to gluten-free flour did not decrease the concentration of starch digestion products. These observations indicate that removal of gluten from wheat flour results into increased amyolytic digestion *in vitro*.

Such studies may be used to identify the foods of potential use for inclusion in diets of diabetic patients²⁷. High protein starchy foods such as legumes are especially useful in diabetic diet²⁸ as they show reduced digestibility²⁹ and lower glycemic response compared to other cereals whose protein content is nearly half that of legumes.

Possibly the wheat flour contains granules with starch molecule in central core surrounded by protein net-work and this protein net-work may inhibit the action of enzyme on starch in the gastrointestinal tract. Possibly the same type of natural starch-protein interaction may account for decreased glycemic response of legumes. Although, the presence of such protein coated starch in legumes is not yet confirmed protein isolates of legumes are found to be in close association with carbohydrates³⁰.

Obese adolescents kept on hypocaloric diet show inter-relationship between glucose and nitrogen balance. It is observed that in such cases with added glucose nitrogen balance becomes significantly more positive. Added dietary carbohydrate stimulates pancreas for insulin secretion which in turn stimulates protein biosynthesis³¹ and inhibits net protein catabolism from skeletal muscle.³²

The protein requirement of an individual varies according to age, sex, and physiological conditions. Normally the protein content in the diet is 1 g /Kg body weight. In India, the recommended dietary allowance is 55 g./day for male and 45 g./day in females. In young children there is an increased need for proteins and this must be kept in view while preparing a dietary plan for juvenile diabetes. In a diet plan of juvenile diabetes, protein requirement is 72 g./day. For an obese diabetic, the daily protein requirement is 40 g. A complete diet plan for the juvenile diabetics and for maturity on-set type of diabetes is as follow:

Diet Plan

Constituents (in g./day)	Juvenile diabetes	Maturity on-set diabetes
Proteins	72	40
Fat	57	33
Carbohydrates	310	135

Summary

Dietary protein deficiency causes altered carbohydrate metabolism in children, adult humans and animals. The decreased insulin release in such malnourished condition results in a condition similar to diabetes mellitus. Lowered protein/DNA ratio in many tissues may be related to lowered capacity for insulin secretion. The effect of natural starch-protein interaction of high protein starchy foods such as legumes are especially useful in diabetic diet as they show reduced digestibility and lowered glycemic response. Inter-relationship of glucose and protein in obese adolescents during hypocalorie diet therapy shows that nitrogen balance is significantly more positive with added glucose. The protein requirement under different states of diabetes mellitus shows that the juvenile diabetes has an increased requirement for proteins, whereas, the maturity onset type has a decreased requirement for proteins

References

1. WHO study group-Diabetes Mellitus, World Health Organization (1985);: 20-25 (Technical report series, 727)
2. Rao R.H., (1984). The role of undernutrition in the pathogenesis of Diabetes Mellitus; *Diabetes Care*; 7: 595-601.
3. Heard C.R.C. (1966). Effect of Severe Protein Caloric deficiency on the endocrine control of carbohydrate metabolism; *Diabetes*, 15: 78-79,
4. Heard C R.C., Turner M.R. (1967). Glucose Tolerance and related factors. in dogs fed diets of suboptimal protein value; *Diabetes*; 16: 96-107.
5. Weinkove C; Weinkove E.A., Pimstone B.L. (1976). Glucose Tolerance and insulin release in malnourished rats; *Clin. Sci. Mol. Med*; 50: 153-63.
6. Younoszai R., Dixit P.K. (1980). Decreased insulin secretion by isolated pancreatic islets from rats fed 4% protein diet, *Proc. Soc. Exp. Biol. Med*; 164: 317-21.

7. Levine L.S., Wright P.O., Marcus F. (1983). Failure to secrete immunoactive insulin by rats fed a low protein diet; *102*: 240-45.
8. Swenne C.J., Grace and Milner R.D.G. (1987). Persistent impairment of insulin secretory response to glucose in adult rats after limited period of protein malnutrition early in life; *Diabetes*; 36: 4: 454-58.
9. Dixit P.K. and Sorenson R.L. (1987). Effect of protein malnutrition on insulin secretion; *Ind. J. Med. Res.* 86: 663-70.
10. Kissane J.M., Robins E. (1954). The fluorometric measurement of deoxyribonucleic acid in animal tissue with special references to the central nervous system; *J. Biochem.*, 233: 184 88.
11. Hinegardner R.T., (1971). An improved fluorometric assay for D.N.A.; *Anat. Biochem*, 39: 197-201.
12. Bradford M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding; *Anat. Biochem.* 72: 248-54.
13. Cheek D.B., Hill H.E., Cornado A., Graham G.G. (1970). Malnutrition in infancy, changes in muscles and adipose tissue before and after rehabilitation; *Paediatr. Res.* 4: 125-44.
14. Cheek D.B., Hill H.E., (1970). Muscle and liver cell growth; Role of hormones and nutritional factors; *Fed. Proc.* 29: 1503-509.
15. Greystone I.E., Cheek D.B. (1969). The effects of reduced caloric intake and increased insulin induced caloric intake on the cell growth of muscle, liver and cerebrum and on skeletal collagen in the post weaning rat; *Paediatr. Res.* 3: 66-76.
16. Trowell H.C., Davis J.N P., Dean R.F.A., London; (1954). Kwashiorkor; *Arnold*, 122- 160.
17. Volk B.W., Lazarus S.S., (1960). Rabbit pancreas in protein malnutrition (experimental Kwashiorkor) and after cortisone administration; *Am. J. Pathol*, 37: 121-35.
18. Platt B.S., Stewart R.J.C. (1967). Experimental protein calorie deficiency; *J. Endocrinol*, 38: 121-43.
19. Madik, Jervis H.R., Anderson P.R., Zimmerman M.R. (1970). A Protein deficient diet; Effect on liver, pancreas, stomach and small intestine of the rat; *Arch. Pathol*, 89: 38-52.
20. Weinkove C., Weinkove E., Timme A., Pimstone B. (1977). Pancreatic islets in malnourished rats-quantitative, histologic and electron microscopic findings; *Arch. Pathol. Lab. Med.* 101: 266-69.

21. Rao A. V., Jenkins D.J.A. (1987). The effect of starch protein interaction in wheat and the glycemic response and rate of *In vitro* digestion; Am. J. Clin. Nutr. 45: 946-52.
22. Crape P.A., Rearen G.M., Olefsky J. (1977). Post prandial plasma glucose and insulin responses to different complex carbohydrates; Diabetes, 28:1178-83.
23. Liener I.E. (1969). Miscellaneous toxic factors. Toxic constituents in plant food stuffs, Acad. Press 430-47.
24. Jaffe W I.G., Lette C.L.V. (1968). Heat labile growth inhibitor factors in beans (*Phaseolus vulgaris*) J. Nutr, 94: 203-10.
25. Yoon J.H., Thompson L.U., Jenkins D.J.A. (1983). The effect of phytic acid *In Vitro* rats of starch digestibility and blood-glucose response; Am. J. Clin. Nutr. 38: 835-42.
26. Hoft C., Derose, Danme J. (1969). Celiac disease in a diabetic child; Lancet; 2: 16 (Letter)
27. Jenkins D.J.A., Wolever T.M.S. (1984). Thorne M.J. The relationship between glycemic response, digestibility and factors influencing the dietary habits of diabetics; Am. J. Clin. Nutr. 40: 1175-91.
28. Anderson J. W., Ward K. (1979). High carbohydrate, high fibre diets for insulin treated men with diabetes mellitus; Am. J. clin. Nubr. 32: 2312-21.
29. Jenkin D.J.A., Wolever T M.S., Taylor R.H. (1980). Rate of digestion of foods and post prandial glycemia in normal and diabetic subjects; Br. Med. J. 2: 7-14.
30. Alli I; Baker R.E. (1980). Constituents of leguminous seeds; the microscopic structure of proteins isolated from phaseolus beans, J. Sc. food. Agr. 31: 1316-22.
31. Sims A.J.W. (1979). Glucose promotes whole body synthesis from infused amino acids in fasting; Lancet; 1: 68-71.
32. Pozefsky T., Felig P., Tobin J.D., Soeldners, J.S.Cahill G.F. (1969). Amino acid balance across tissues of the forearm in post-absorptive man; Effect of insulin at two dose levels J. Clin. Invest; 48: 2273-82.